AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit

IVD

For Professional Use Only

Instruction Manual

KEY TO SYMBOLS USED

REF	Catalogue number		Caution
LOT	Batch code		Sufficient for
IVD	*In vitro* diagnostic medical device		Use-by Date
VER	Version		Consult instructions for use
	Temperature limit		Keep away from sunlight
	Manufacturer	NCA	Negative control of amplification
	Date of manufacture	C–	Negative control of extraction
EC REP	Authorized representative in the European Community	C+	Positive control of amplification
		IC	Internal control

1. INTENDED USE

AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit is an *in vitro* nucleic acid amplification test for simultaneous detection of *Trichomonas vaginalis* and *Neisseria gonorrhoeae* DNA in the clinical material (urogenital, rectal, and oropharyngeal swabs; conjunctival discharge; urine samples, prostate gland secretion) using real-time hybridization-fluorescence detection of amplified products.

NOTE: The results of PCR analysis are taken into account in complex diagnostics of disease.

2. PRINCIPLE OF PCR DETECTION

T.vaginalis / N.gonorrhoeae detection by the polymerase chain reaction (PCR) is based on the amplification of the pathogen genome specific region using specific *T.vaginalis / N.gonorrhoeae* primers. In the real-time PCR, the amplified product is detected with the use of fluorescent dyes. These dyes are linked to oligonucleotide probes which bind specifically to the amplified product during thermocycling. The real-time monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run.

AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit is a qualitative test that contains the Internal Control (Internal Control-FL (IC)). It must be used in the extraction procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition.

AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit uses "hot-start," which greatly reduces the frequency of nonspecifically primed reactions. "Hot-start" is guaranteed by using chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

The PCR kit contains the system for prevention of contamination by amplicons using the enzyme uracil-DNA-glycosylase (UDG) and dUTP. The enzyme UDG recognizes and catalyzes the destruction of the DNA containing deoxyuridine, but has no effect on DNA containing deoxythymidine. Deoxyuridine is absent in the authentic DNA, but is always present in amplicons, because dUTP is a part of dNTP mixture in the reagents for the amplification. Due to the deoxyuridine containing contaminating amplicons are sensitive to the destruction by UDG before the DNA-target amplification. So the amplicons cannot be amplified.

The enzyme UDG is thermolabile. It is inactivated by heating at temperature above 50 °C. Therefore, UDG does not destroy the target amplicons which are accumulated during PCR.

Table 1

Channel for fluorophore	FAM	JOE	ROX
DNA-target	*Trichomonas vaginalis*	*Neisseria gonorrhoeae*	Internal Control-FL (IC) DNA
Target gene	Trichomonas vaginalis repeated DNA target for PCR identification	*gene* 16S rRNA	Artificially synthesized sequence

3. CONTENT

AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit is produced in 1 form:

variant FRT-100 F, REF R-B65-F(RG,iQ)-CE.

Variant FRT-100 F includes:

Reagent	Description	Volume, ml	Quantity
PCR-mix-1-FL *T.vaginalis / N.gonorrhoeae*	clear liquid from colorless to light lilac colour	1.2	1 tube
PCR-mix-2-FRT	colorless clear liquid	0.3	2 tubes
Polymerase (TaqF)	colorless clear liquid	0.03	2 tube
Positive Control complex (C+)	colorless clear liquid	0.2	1 tube
DNA-buffer	colorless clear liquid	0.5	1 tube
Negative Control (C–)*	colorless clear liquid	1.2	1 tube
Internal Control-FL (IC)**	colorless clear liquid	1.0	1 tube

* must be used in the extraction procedure as Negative Control of Extraction.

** add **10 µl** of **Internal Control-FL (IC)** during the DNA extraction procedure directly to the sample/lysis mixture (see the **DNA-sorb-AM** REF K1-12-100-CE protocol).

Variant FRT-100 F is intended for 110 reactions (including controls).

4. ADDITIONAL REQUIREMENTS

- DNA extraction kit.
- Transport medium.
- Disposable powder-free gloves and laboratory coat.
- Pipettes (adjustable).
- Sterile pipette tips with aerosol filters (up to 100 µl).
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with a rotor for 2 ml reaction tubes.
- PCR box.
- Real-time instruments (for example, Rotor-Gene 3000/6000 (Corbett Research, Australia); Rotor-Gene Q (QIAGEN, Germany); iCycler iQ5 (Bio-Rad, USA); Mx3000P (Stratagene, USA).
- Disposable polypropylene tubes:
  a) thin-walled 0.2-ml PCR tubes with domed caps if a plate-type instrument is used;
  b) thin-walled 0.2-ml PCR tubes with flat caps or strips of four 0.1-ml Rotor-Gene PCR tubes if a rotor-type instrument is used.
- Refrigerator for 2–8 °C.
- Deep-freezer at the temperature from minus 24 to minus 16 °C.
- Reservoir for used tips.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol filters and use a new tip for every procedure.
- Store all extracted positive material (specimens, controls and amplicons) away from all other reagents and add it to the reaction mix in a distantly separated facility.
- Thaw all components thoroughly at room temperature before starting an assay.
- When thawed, mix the components and centrifuge briefly.
- Use disposable protective gloves and laboratory cloths, and protect eyes while samples and reagents handling. Thoroughly wash hands afterwards.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all specimens and unused reagents in accordance with local regulations.
- Samples should be considered potentially infectious and handled in biological cabinet in compliance with appropriate biosafety practices.
- Clean and disinfect all samples or reagents spills using a disinfectant, such as 0.5 % sodium hypochlorite or another suitable disinfectant.
- Avoid inhalation of vapors, samples and reagents contact with the skin, eyes, and mucous membranes. Harmful if swallowed. If these solutions come into contact, rinse the injured area immediately with water and seek medical advice if necessary.
- Safety Data Sheets (SDS) are available on request.
- Use of this product should be limited to personnel trained in DNA amplification techniques.
- Workflow in the laboratory must be one-directional, beginning in the Extraction Area and moving to the Amplification and Detection Area. Do not return samples, equipment and reagents in the area where the previous step was performed.

⚠ Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

6. SAMPLING AND HANDLING

NOTE: Obtaining samples of biological materials for PCR-analysis, transportation and storage are described in the manufacturer's handbook [1]. It is recommended that this handbook is read before starting work.

AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit is intended for analysis of the DNA extracted with DNA extraction kits from the clinical material (urogenital, rectal, and oropharyngeal swabs; conjunctival discharge; urine samples (a sediment of the first portion of the morning specimen); prostate gland secretion).

7. WORKING CONDITIONS

AmpliSens® *T.vaginalis / N.gonorrhoeae-MULTIPRIME*-FRT PCR kit should be used at 18–25 °C.

8. PROTOCOL

8.1. DNA Extraction

It is recommended to use the following nucleic acid extraction kits:

- **DNA-sorb-AM**, REF K1-12-100-CE;
- For other nucleic acid extraction kits see Guidelines [2].

The DNA extraction of each test sample is carried out in the presence of **Internal Control-FL (IC).**

In the extraction procedure it is necessary to carry out the control reactions as follows:

C– – Add **100 µl of Negative Control (C–)** to the tube labeled C– (Negative Control of Extraction).

NOTE: Extract DNA according to the manufacturer's protocol.

8.2. Preparing PCR

The type of tubes depends on the PCR instrument used for analysis.
Use disposable filter tips for adding reagents, DNA and control samples into tubes.

8.2.1 Preparing tubes for PCR

The total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.

1. Thaw the tube with **PCR-mix-2-FRT**. Vortex the tubes with **PCR-mix-1-FL *T.vaginalis / N.gonorrhoeae*, PCR-mix-2-FRT,** and **polymerase (TaqF)** and then centrifuge briefly. Take the required number of tubes/strips for amplification of the DNA obtained from clinical and control samples.
2. For N reactions (including 2 controls) add to a new tube:
   10·(N+1) µl of **PCR-mix-1-FL *T.vaginalis / N.gonorrhoeae***,
   5.0·(N+1) µl of **PCR-mix-2-FRT**,
   0.5·(N+1) µl of **polymerase (TaqF)**.

Vortex the tube, then centrifuge it briefly. Transfer **15 µl** of the prepared mixture to each tube.

3 Using tips with aerosol filter, add **10 µl** of **DNA samples** obtained at the DNA extraction stage.
4 Carry out the control amplification reactions:
NCA – Add **10 µl** of **DNA-buffer** to the tube labeled NCA (Negative Control of Amplification).
C+ – Add **10 µl** of **Positive Control complex** to the tube labeled C+ (Positive control of Amplification).
C– – Add **10 µl** of **the sample extracted from the Negative Control (C–) reagent** to the tube labeled C– (Negative control of Extraction).

8.2.2. Amplification

1. Create a temperature profile on your instrument as follows:

Table 2

AmpliSens-1 amplification program

	Rotor-type Instruments[1]			Plate-type Instruments[2]		
Step	**Temperature, °C**	**Time**	**Cycles**	**Temperature, °C**	**Time**	**Cycles**
Hold	**95**	**15 min**	**1**	**95**	**15 min**	**1**
Cycling	**95**	**5 s**	**5**	**95**	**5 s**	**5**
	60	**20 s**		**60**	**20 s**	
	72	**15 s**		**72**	**15 s**	
Cycling 2	**95**	**5 s**	**40**	**95**	**5 s**	**40**
	60	**20 s Fluorescence acquiring**		**60**	**30 s Fluorescence acquiring**	
	72	**15 s**		**72**	**15 s**	

Fluorescent signal is detected in the channels designed for the FAM, JOE and ROX fluorophores (other channels are enabled if several tests are simultaneously carried out in a single run).

2. Adjust the fluorescence channel sensitivity according to the *Important Product Information Bulletin* and Guidelines [2].
3. Insert tubes into the reaction module of the device.
4. Run the amplification program with fluorescence detection.
5. Analyze results after the amplification program is completed.

9. DATA ANALYSIS

Analysis of results is performed by the software of the real-time PCR instrument used by measuring fluorescence signal accumulation in three channels:

– The signal of the *Trichomonas vaginalis* DNA amplification product is detected in the channel for the FAM fluorophore.
– The signal of the *Neisseria gonorrhoeae* DNA amplification product is detected in the channel for the JOE fluorophore.
– The signal of the IC DNA amplification product is detected in the channel for the ROX fluorophore.

Results are interpreted by the crossing (or not-crossing) the fluorescence curve with the threshold line set at the specific level that corresponds to the presence (or absence) of a *Ct* value of the DNA sample in the corresponding column of the results grid.

Principle of interpretation is the following:

– ***Trichomonas vaginalis*** DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the FAM fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
– ***Neisseria gonorrhoeae*** DNA is **detected** if the *Ct* value is determined in the results grid in the channel for the JOE fluorophore. Moreover, the fluorescence curve of the sample should cross the threshold line in the area of typical exponential growth of fluorescence.
– ***Trichomonas vaginalis and Neisseria gonorrhoeae*** DNA are **not detected** in a sample if the *Ct* value is not determined (absent) in the channels for FAM and JOE fluorophores, whereas the *Ct* value determined in the channel for the ROX fluorophore is less than the boundary *Ct* value specified in the Important Product Information Bulletin.
– The result is **invalid** if the *Ct* value is not determined (absent) in the channels for FAM, JOE or ROX fluorophores. In such cases, the PCR analysis should be repeated.

NOTE: Boundary *Ct* values are specified in the *Important Product Information Bulletin* enclosed to the PCR kit. See also Guidelines [2].

The result of the analysis is considered reliable only if the results obtained for the Positive and Negative Controls of amplification as well as for the Negative Control of extraction are correct (see Table 3).

Table 3

Results for controls

Control	Stage for control	*Ct* value in the channel for fluorophore	
		FAM, JOE	**ROX**
C–	DNA extraction	Absent	< boundary value
NCA	PCR	Absent	Absent
C+	PCR	< boundary value	< boundary value

10. TROUBLESHOOTING

Results of analysis are not taken into account in the following cases:

1. If the *Ct* value determined for the Positive Control of Amplification (C+) in the channels for the FAM and\or JOE and\or ROX fluorophores is greater than the boundary *Ct* value or absent, the amplification should be repeated for all samples in which the boundary *Ct* is absent in the channels for the FAM and\or JOE and\or ROX fluorophores.
2. If the *Ct* value is determined for the Negative Control of Amplification (NCA) and/or Negative Control of Extraction (C–) in the channels for the FAM and/or JOE fluorophores, the PCR analysis should be repeated for all samples in which the *Ct* value is determined in the channels for the FAM and/or JOE fluorophores..

If you have any further questions or if encounter problems, please contact our Authorized representative in the European Community.

11. TRANSPORTATION

AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT PCR kit should be transported at 2–8 °C for no longer than 5 days

12. STABILITY AND STORAGE

All components of the **AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT** PCR kit are to be stored at 2–8 °C when not in use (except for polymerase (TaqF) and PCR-mix-2-FRT). All components of the **AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT** PCR kit are stable until the expiry date on the label. The shelf life of reagents before and after the first use is the same, unless otherwise stated.

NOTE: Polymerase (TaqF) and PCR-mix-2-FRT are to be stored at the temperature from minus 24 to minus 16 °C when not in use.

NOTE: PCR-mix-1-FL *T.vaginalis / N.gonorrhoeae* is to be kept away from light.

13. SPECIFICATIONS

13.1. Sensitivity

The analytical sensitivity of **AmpliSens® *T.vaginalis / N.gonorrhoeae*-MULTIPRIME-FRT** PCR kit is the following:

Clinical material	Transport medium	Nucleic acid extraction kit	Microorganism	Sensitivity, GE/ml[3]
Urogenital swabs	**Transport medium for swabs** (REF 956-CE, REF 987-CE) or **Transport medium with mucolytic agent** (REF 952-CE, REF 953-CE	DNA-sorb-AM	*Trichomonas vaginalis*	$5x10^2$
			Neisseria gonorrhoeae	$5x10^2$
Urine[4]	—	DNA-sorb-AM	*Trichomonas vaginalis*	10^3
			Neisseria gonorrhoeae	10^3

NOTE: The analytical sensitivity of each microorganism does not change even at high concentrations of the other microorganism (to 10^9 GE/ml).

[1] For example, Rotor-Gene 3000, Rotor-Gene 6000, Rotor-Gene Q.
[2] For example, iQ5, Mx3000P, Mx3000.
[3] The quantity of genome equivalents of microorganism per 1 ml of the sample.
[4] Treatment is needed.

13.2. Specificity

The analytical specificity of **AmpliSens® *T.vaginalis* / *N.gonorrhoeae*-MULTIPRIME-FRT** PCR kit is ensured by the selection of specific primers and probes as well as strict reaction conditions. The primers and probes have been checked for possible homologies to all sequences published in gene banks by sequence comparison analysis.

Nonspecific responses were absent while testing human DNA samples as well as a DNA panel of the following microorganisms: *Gardnerella vaginalis, Lactobacillus* spp., *Escherichia coli, Staphylococcus* spp., *Streptococcus* spp., *Candida albicans, Mycoplasma hominis, Ureaplasma urealyticum, Ureaplasma parvum, Mycoplasma genitalium, Chlamydia trachomatis, Neisseria* spp., *Neisseria gonorrhoeae, Trichomonas vaginalis, Treponema pallidum, Toxoplasma gondii, HSV* types 1 and 2, *CMV,* and *HPV*.

The clinical specificity of **AmpliSens® *T.vaginalis* / *N.gonorrhoeae*-MULTIPRIME-FRT** PCR kit was confirmed in laboratory clinical trials.

14. REFERENCES

1. Handbook “Sampling, Transportation, and Storage of Clinical Material for PCR Diagnostics”, developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology” of Federal Service for Surveillance on Consumers’ Rights Protection and Human Well-Being.
2. Guidelines “Real-Time PCR Detection of STIs and Other Reproductive Tract Infections”, developed by Federal Budget Institute of Science “Central Research Institute for Epidemiology” of Federal Service for Surveillance on Consumers’ Rights Protection and Human Well-Being.

15. QUALITY CONTROL

In compliance with Federal Budget Institute of Science “Central Research Institute for Epidemiology” ISO 13485-Certified Quality Management System, each lot of **AmpliSens® *T.vaginalis* / *N.gonorrhoeae*-MULTIPRIME-FRT** PCR kit has been tested against predetermined specifications to ensure consistent product quality.

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
29.06.11 LA	Cover page, text	The name of Institution was changed to Federal Budget Institution of Science “Central Research Institute for Epidemiology”
17.02.16 PM	Through the text	Corrections in accordance with the template
	1. Intended use	The clinical material was specified
	8.1. DNA extraction	Information about controls of extraction was added
	9. Data analysis	The sections were rewritten
	10. Troubleshooting	
15.03.18 PM	Footer, 3. Content	REF R-B65(iQ)-CE was deleted
10.09.18 EM	3. Content	The colour of the reagent was specified
17.01.19 PM	2. Principle of PCR detection	The information about the enzyme UDG was added. The information about “hot-start” was corrected
29.04.20 KK	Through the text	The text formatting was changed
	2. Principle of PCR detection	The table with targets was added.
	Footer	The phrase “Not for use in the Russian Federation” was added
30.10.20 MA	Through the text, Footer	The information about variant FRT REF R-B65(RG)-CE was deleted
18.03.21 MM	—	The name, address and contact information for Authorized representative in the European Community was changed

AmpliSens®

EC REP

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