



For *in Vitro* Diagnostic Use

TABLE OF CONTENTS

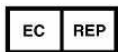
1. INTENDED USE	3
2. PRINCIPLE OF PCR DETECTION	3
3. CONTENT	3
4. ADDITIONAL REQUIREMENTS	5
5. GENERAL PRECAUTIONS	6
6. SAMPLING AND HANDLING	6
7. PROTOCOL	7
8. DATA ANALYSIS	9
9. TROUBLESHOOTING	10
10. STABILITY AND STORAGE	11
11. SPECIFICATIONS	11
12. REFERENCES	12
13. QUALITY CONTROL	12
14. EXPLANATION OF SYMBOLS	13

AmpliSens[®] ARVI-screen-FRT

PCR kit

Instruction Manual

AmpliSens[®]



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1. INTENDED USE

AmpliSens® ARVI-screen-FRT PCR kit is an in vitro nucleic acid amplification test for multiplex detection and identification of specific nucleic acid fragments of pathogens that cause acute respiratory viral infections – *human Respiratory Syncytial virus (hRSV)* RNA; *human Metapneumovirus (hMPv)* RNA; *human Parainfluenza virus-1-4 (hPiv)* RNA; OC43, E229, NL63, and HKUI *human Coronavirus (hCov)* RNA; *human Rhinovirus (hRv)* RNA; *human B, C, and E Adenovirus (hAdv)* DNA; and *human Bocavirus (hBov)* DNA – in the clinical material (nasal and oropharyngeal swabs, sputum, aspirate of trachea, bronchoalveolar lavage, bronchial washing fluid, and autopsy material) by using real-time hybridization-fluorescence detection.

2. PRINCIPLE OF PCR DETECTION

ARVI detection by the polymerase chain reaction (PCR) is based on the amplification of the pathogen genome specific region using specific ARVI primers. In real-time PCR, the amplified product is detected using fluorescent dyes. These dyes are linked to oligonucleotide probes which bind specifically to the amplified product during thermocycling. The real-time PCR monitoring of fluorescence intensities during the real-time PCR allows the detection of accumulating product without re-opening the reaction tubes after the PCR run. **AmpliSens® ARVI-screen-FRT** PCR kit is a qualitative test that contains the Internal Control (IC). It must be used in the extraction procedure in order to control the extraction process of each individual sample and to identify possible reaction inhibition. **AmpliSens® ARVI-screen-FRT** PCR kit uses “hot-start,” which greatly reduces the frequency of nonspecifically primed reactions. “Hot-start” is guaranteed by separation of nucleotides and Taq-polymerase by using a chemically modified polymerase (TaqF). The chemically modified polymerase (TaqF) is activated by heating at 95 °C for 15 min.

3. CONTENT

AmpliSens® ARVI-screen-FRT PCR kit is produced in 3 forms:

AmpliSens® ARVI-screen-FRT PCR kit variant FRT (for use with RG)

REF R-V57(RG)-CE;

AmpliSens® ARVI-screen-FRT PCR kit variant FRT (for use with iQ, Dt)

REF R-V57(iQ,Dt)-CE;

AmpliSens® ARVI-screen-FRT PCR kit variant FRT-100 F (for use with RG, iQ, Dt)

REF R-V57-100-F(RG,iQ,Dt)-CE.

AmpliSens® ARVI-screen-FRT PCR kit variant FRT includes:

Reagent	Description	Volume (ml)	Amount
PCR-mix-1-FL hRSv – hMPv ready-to-use single-dose test tubes (under wax)	colorless clear liquid	0.008	55 tubes of 0.2 ml
PCR-mix-1-FL hPiv 1/3 ready-to-use single-dose test tubes (under wax)	colorless clear liquid	0.008	55 tubes of 0.2 ml
PCR-mix-1-FL hPiv 2/4 ready-to-use single-dose test tubes (under wax)	colorless clear liquid	0.008	55 tubes of 0.2 ml
PCR-mix-1-FL hCov ready-to-use single-dose test tubes (under wax)	colorless clear liquid	0.008	55 tubes of 0.2 ml
PCR-mix-1-FL hAdv – hBov ready-to-use single-dose test tubes (under wax)	colorless clear liquid	0.008	55 tubes of 0.2 ml
PCR-mix-1-FL hRv ready-to-use single-dose test tubes (under wax)	colorless clear liquid	0.008	55 tubes of 0.2 ml
PCR-mix-2-FL	colorless clear liquid	0.77	4 tubes
Positive Control cDNA hRSv - hMPv (C ⁺ _{hRSv-hMPv})	colorless clear liquid	0.1	1 tube
Positive Control cDNA hPiv 1/3 (C ⁺ _{hPiv 1/3})	colorless clear liquid	0.1	1 tube
Positive Control cDNA hPiv 2/4 (C ⁺ _{hPiv 2/4})	colorless clear liquid	0.1	1 tube
Positive Control cDNA hRv (C ⁺ _{hRv})	colorless clear liquid	0.1	1 tube
Positive Control cDNA hCov (C ⁺ _{hCov})	colorless clear liquid	0.1	1 tube
Positive Control DNA hAdv - hBov (C ⁺ _{hAdv-hBov})	colorless clear liquid	0.1	1 tube
Positive Control STI-88 (CS+)	colorless clear liquid	0.1	3 tubes
TE-buffer	colorless clear liquid	0.5	1 tube
Negative Control (C-)*	colorless clear liquid	1.2	1 tube
Internal Control STI-rec (IC)**	colorless clear liquid	0.12	5 tubes

* must be used in the extraction procedure as Negative Control of Extraction.

** add 10 µl of Internal Control during the DNA extraction procedure directly to the sample/lysis mixture (see RIBO-sorb **REF** K2-1-Et-100-CE and RIBO-prep **REF** K2-9-Et-100-CE protocols).

AmpliSens® ARVI-screen-FRT PCR kit variant FRT is intended for 55 reactions for every PCR-mix-1-FL (including controls).

AmpliSens® ARVI-screen-FRT PCR kit variant FRT-100 F includes:

Reagent	Description	Volume, ml	Amount
PCR-mix-1-FL-F hRSv – hMpv	colorless clear liquid	0.2	5 tubes
PCR-mix-1-FL-F hPiv 1/3	colorless clear liquid	0.2	5 tubes
PCR-mix-1-FL-F hPiv 2/4	colorless clear liquid	0.2	5 tubes
PCR-mix-1-FL-F hCov	colorless clear liquid	0.2	5 tubes
PCR-mix-1-FL-F hAdv – hBov	colorless clear liquid	0.2	5 tubes
PCR-mix-1-FL-F hRv	colorless clear liquid	0.2	5 tubes
PCR-mix-2-FRT	colorless clear liquid	0.6	6 tubes
Polymerase (TaqF)	colorless clear liquid	0.06	6 tubes
Positive Control cDNA hRSv – hMpv (C ⁺ _{hRSv-hMpv})	colorless clear liquid	0.1	2 tubes
Positive Control cDNA hPiv 1/3 (C ⁺ _{hPiv 1/3})	colorless clear liquid	0.1	2 tubes
Positive Control cDNA hPiv 2/4 (C ⁺ _{hPiv 2/4})	colorless clear liquid	0.1	2 tubes
Positive Control cDNA hRv (C ⁺ _{hRv})	colorless clear liquid	0.1	2 tubes
Positive Control cDNA hCov (C ⁺ _{hCov})	colorless clear liquid	0.1	2 tubes
Positive Control DNA hAdv – hBov (C ⁺ _{hAdv-hBov})	colorless clear liquid	0.1	2 tubes
Positive Control STI-88 (CS+)	colorless clear liquid	0.1	6 tubes
TE-buffer	colorless clear liquid	0.5	2 tubes
Negative Control (C-)*	colorless clear liquid	1.2	2 tubes
Internal Control STI-rec (IC)**	colorless clear liquid	0.12	10 tubes

* must be used in the extraction procedure as Negative Control of Extraction.

** add 10 µl of Internal Control during the DNA extraction procedure directly to the sample/lysis mixture (see RIBO-sorb [REF](#) K2-1-Et-100-CE and RIBO-prep [REF](#) K2-9-Et-100-CE protocols).

AmpliSens® ARVI-screen-FRT PCR kit variant FRT-100 F is intended for 100 reactions for every PCR-mix-1-FL (including controls).

4. ADDITIONAL REQUIREMENTS

- DNA/RNA extraction kit.
- Reverse transcription kit.
- Transport medium.
- Disposable powder-free gloves and laboratory coat.
- Pipettes (adjustable).

- Sterile pipette tips with aerosol barriers (up to 200 µl).
- Tube racks.
- Vortex mixer.
- Desktop centrifuge with rotor for 2-ml reaction tubes.
- PCR box.
- Personal thermocyclers (for example, Rotor-Gene 6000 (Corbett Research, Australia), Rotor-Gene Q (Qiagen, Germany), iCycler iQ or iQ5 (Bio-Rad, USA), or equivalent).
- Disposable polypropylene microtubes for PCR (0.2-ml or 0.1-ml); for example, Axygen, USA; Corbett Research, Australia; Qiagen, Germany).
- Refrigerator for temperature 2-8 °C.
- Deep-freezer with temperature ≤16 °C.
- Waste bin for used tips.

5. GENERAL PRECAUTIONS

The user should always pay attention to the following:

- Use sterile pipette tips with aerosol barriers and use new tip for every procedure.
- Store and handle amplicons away from all other reagents.
- Thaw all components thoroughly at room temperature before starting detection.
- When thawed, mix the components and centrifuge briefly.
- Use disposable gloves, laboratory coats, protect eyes while samples and reagents handling. Thoroughly wash hands afterward.
- Do not eat, drink, smoke, apply cosmetics, or handle contact lenses in laboratory work areas.
- Do not use a kit after its expiration date.
- Dispose of all samples and unused reagents in compliance with local authorities requirements.
- Samples should be considered potentially infectious and handled in a biological cabinet in accordance with appropriate biosafety practices.
- Clean and disinfect all sample or reagent spills using a disinfectant such as 0.5 % sodium hypochlorite, or other suitable disinfectant.
- Avoid contact with the skin, eyes and mucosa. If skin, eyes and mucosa contact, immediately flush with water, seek medical attention.
- Material Safety Data Sheets (MSDS) are available on request.
- Use of this product should be limited to personnel trained in the techniques of DNA amplification.
- The laboratory process must be one-directional, it should begin in the Extraction Area and then move to the Amplification and Detection Area. Do not return samples, equipment and reagents to the area in which the previous step was performed.



Some components of this kit contain sodium azide as a preservative. Do not use metal tubing for reagent transfer.

6. SAMPLING AND HANDLING



Obtaining samples of biological materials for PCR-analysis, transportation and storage is described in manufacturer's handbook [1]. It is recommended that this handbook is read before starting work.

AmpliSens® ARVI-screen-FRT PCR kit is intended for analysis of DNA/RNA extracted from:

- nasal and oropharyngeal swabs;
- sputum (or aspirate of trachea or throat);

- bronchoalveolar lavage or bronchial washing fluid;
- autopsy material.

7. PROTOCOL

Complete analysis includes following steps:

- DNA/RNA extraction from clinical samples.
- Reverse transcription.
- Amplification and real-time hybridization-fluorescence detection.
- Data analysis and interpretation of results.

7.1 DNA/RNA extraction

It is recommended to use following nucleic acid extraction kits:

- RIBO-sorb, **REF** K2-1-Et-100-CE.
- RIBO-prep, **REF** K2-9-Et-100-CE.

DNA/RNA extraction from every clinical sample is carried out in presence of **IC STI-rec**.

7.2 Reverse transcription

It is recommended to use following RT reagents kits for complementary DNA (cDNA) synthesis from RNA.

- REVERTA-L, **REF** K3-4-50-CE, which contains RT-G-mix-1.



Carry out the reverse transcription procedure according to the manufacturer's instruction.

7.3 Preparing PCR

The total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.



At the amplification step, positive controls (see Table 1), CS+, and NCA are used in every experiment in order to control reagent purity and carefulness of operator's work. C– is also tested at the amplification step.

Table 1

Compliance of names of PCR-mixes-1-FL and positive controls of ARVI pathogens

PCR-mix-1-FL	Positive control samples (C+)
<i>hRSv - hMpv</i>	Positive Control cDNA <i>hRSv - hMpv</i>
<i>hAdv - hBov</i>	Positive Control DNA <i>hAdv - hBov</i>
<i>hRv</i>	Positive Control cDNA <i>hRv</i>
<i>hPiv 1/3</i>	Positive Control cDNA <i>hPiv 1/3</i>
<i>hPiv 2/4</i>	Positive Control cDNA <i>hPiv 2/4</i>
<i>hCov</i>	Positive Control cDNA <i>hCov</i>

7.3.1 Preparing tubes for PCR

The type of tubes depends on the type of PCR real-time instrument.

Use disposable tips with aerosol barriers for adding reagents, cDNA and control samples into tubes.

Variant FRT-100 F

The total reaction volume is **25 µl**, the volume of DNA sample is **10 µl**.

1. Thaw the tubes with PCR-mixes-1-FL-F.
2. For carrying out N reactions (including 2 controls), mix in a new tube: **10·(N+1) µl of PCR-mix-1-FL-F**, **5.0·(N+1) µl of PCR-mix-2-FRT** and **0.5·(N+1) µl of polymerase (TaqF)**. Vortex the tube, then centrifuge shortly.
3. Transfer **15 µl** of the prepared mixture into each tube.
4. Add **10 µl** of **cDNA** obtained at the reverse transcription stage into the prepared tubes.
5. Carry out the control reactions (for every **PCR-mix-1-FL-F**, see Table 1):

NCA - Add **10 µl** of **TE-buffer** to the tube labeled NCA (Negative Control of Amplification).

C+ - Add **10 µl** of **Positive Control** to tubes labeled C+ (**C+_{hRSv-hMpv}** etc., depending on the PCR-mix-1-FL, see Table 1).

CS+ - Add **10 µl** of **Positive Control STI-88** to the tube labeled CS+.

C– Add **10 µ** of the sample extracted from **Negative Control** to the tube labeled C–.

6. Centrifuge the reaction mixture for 1-2 s.

7.3.2 Amplification

1. Program the PCR instrument (with real-time detection) according to Table 2.

Table 2

ARVI-screen amplification program

Step	Rotor-type instruments ¹			Plate-type instruments ²		
	Temperature, °C	Time	Cycles	Temperature, °C	Time	Cycles
1	95	15 min	1	95	15 min	1
2	95	10 s	10	95	10 s	10
	54	20 s		54	25 s	
	72	10 s		72	25 s	
3	95	10 s	35	95	10 s	35
	54	20 s		54	25 s	
		Fluorescence detection			Fluorescence detection	
72	10 s	72	25 s			

Fluorescence is detected is in FAM/Green, JOE/Yellow/HEX and ROX/Orange fluorescent channels.

¹ For example, Rotor-Gene 3000, Rotor-Gene 6000, Rotor-Gene Q, or equivalent.

² For example, iQ5, iCycler iQ, or equivalent.



It is **not allowed** to perform «*Rhinovirus*» test together with other tests from AmpliSens® ARVI-screen-FRT PCR kit when working with iCycler iQ and iQ5 instruments.

2. Place PCR tubes into the PCR instrument.
3. Run amplification and signal detection program.
4. After measurement, start data analysis and interpretation of results.

8. DATA ANALYSIS

The results are interpreted by the software of Rotor-Gene 3000 or Rotor-Gene 6000 or iCycler iQ or iQ5 instrument by the crossing (or not-crossing) of the fluorescence curve with the threshold line.



Data analysis for each PCR-mix-1 should be performed individually, after withdrawal of tubes corresponding to the PCR-mix-1 used. For the «*Rhinovirus*» (*hRv*) test analysis, it is necessary to use **ONLY FAM** and **ROX** channels.

Table 3

Correspondence of PCR-mixes-1-FL and channels for ARVI pathogen detection

PCR-mix-1-FL	Fluorescence detection		
	FAM/Green	JOE/Yellow/HEX	ROX/Orange
<i>hRSv-hMpv</i>	IC	<i>hRSv</i>	<i>hMpv</i>
<i>hAdv-hBov</i>	IC	<i>hBov</i>	<i>hAdv</i>
<i>hRv</i>	IC	–	<i>hRv</i>
<i>hPiv 1/3</i>	IC	<i>hPiv 3</i>	<i>hPiv 1</i>
<i>hPiv 2/4</i>	IC	<i>hPiv 2</i>	<i>hPiv 4</i>
<i>hCov</i>	IC	<i>NL-63, 229E</i>	<i>HKU-1, OC 43</i>

Principle of interpretation of results:

- DNA/RNA of an ARVI pathogen is **detected** if the Ct value for this sample is determined in the results grid in the corresponding channel. The fluorescence curve for this sample should cross the threshold line in the interval of exponential growth of the fluorescence curve.
- DNA/RNA of an ARVI pathogen is **not detected** if the Ct value for tested sample is not determined (absent) in the results grid in the corresponding channel and if the Ct value in the results grid in the FAM channel does not exceed the specified boundary value.
- Result is considered to be **invalid** if the Ct for the tested sample is not determined (absent) in the corresponding channel for ARVI pathogens (see Table 3) and if the Ct value in the FAM/Green channel is absent or exceeds the specified boundary value. In this case the analysis of the sample should be repeated from the DNA/RNA extraction step.

The results of analysis are considered reliable only if the results obtained for positive and negative controls of amplification and negative control of extraction are correct (see Table 4).

Results for controls

Control	Stage for control	Ct value in channel		
		FAM/Green	JOE/Yellow/HEX	ROX/Orange
		Detection of IC	Detection of ARVI pathogen	Detection of ARVI pathogen
C–	RNA extraction	Pos (< boundary value*)	<u>Neg</u>	<u>Neg</u>
NCA	Amplification	<u>Neg</u>	<u>Neg</u>	<u>Neg</u>
CS+	Amplification	Pos (< boundary value*)	<u>Neg</u>	<u>Neg</u>
C+	Amplification	<u>Neg</u>	Pos (< boundary value**)	Pos (< boundary value*)

* For boundary values, see the Guidelines and Important Product Information Bulletin for AmpliSens® ARVI-screen-FRT PCR kit.

** **Positive Control cDNA *hRv*** is not determined in the JOE channel.

9. TROUBLESHOOTING

- If the Ct value is absent in all channels or the Ct value in the FAM/Green channel is higher than the specified boundary value, PCR should be repeated. If the same result is obtained, the extraction stage for the sample should be repeated. If the IC signal of this sample was detected normally in any other PCR test, it is not necessary to repeat the extraction stage (if iCycler iQ or iQ5 instruments are used).
- If the Ct value is present for C– in the JOE/Yellow/HEX and ROX/Orange channels and/or for NCA in all channels in the results grid, it indicates contamination of reagents or samples. In such cases, the results of analysis must be considered as invalid. Test analysis must be repeated and measures to detect and eliminate the source of contamination must be taken.
- If no signal is detected for the positive controls of amplification, it may suggest that the programming of the temperature profile of the used Instrument was incorrect, or that the configuration of the PCR reaction was incorrect, or that the storage conditions for kit components did not comply with the manufacturer's instruction, or that the reagent kit expired. Programming of the used instrument, storage conditions, and the expiration date of the reagents should be checked, and then PCR should be repeated.
- If a positive result (the fluorescence curve crosses the threshold line) is detected for a sample that has a fluorescence curve without the typical exponential growth phase (the curve is

linear), this may suggest incorrect setting of the threshold line or incorrect calculation of baseline parameters. Such a result should not be considered as positive. Once the threshold line has been set correctly, PCR analysis of the sample should be repeated (if iCycler iQ or iQ5 instruments are used).

10. STABILITY AND STORAGE

All components of **AmpliSens® ARVI-screen-FRT** PCR kit variant FRT-100 F (except for PCR-mix-2-FRT, PCR-mixes-1-FL-F (0.2 ml), and polymerase (TaqF)) are to be stored at 2–8 °C. PCR-mix-2-FRT, PCR-mixes-1-FL-F (0.2 ml), and polymerase (TaqF) are to be stored at ≤ –16 °C.

All components of the **AmpliSens® ARVI-screen-FRT** PCR kit are stable until the expiration date on the label.



PCR-mix-1-FL-F *hRSv* – *hMpv*, PCR-mix-1-FL-F *hPiv* 1/3, PCR-mix-1-FL-F *hPiv* 2/4, PCR-mix-1-FL-F *hCov*, PCR-mix-1-FL-F *hAdv* – *hBov*, and PCR-mix-1-FL-F *hRv* are to be kept away from light.

11. SPECIFICATIONS

11.1 Sensitivity

For samples from nasal and oropharyngeal swabs:

Pathogen	RNA/DNA extraction kit	Amplification and detection kit	Analytical sensitivity, GE/ml ³
<i>hRSv</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	1x10 ³
<i>hMpv</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	1x10 ³
<i>hPiv</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	1x10 ³
<i>hCov</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	1x10 ⁴
<i>hBov</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	1x10 ³
<i>hAdv</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	5x10 ³
<i>hRv</i>	RIBO-sorb, RIBO-prep, NucliSENS easyMAG	AmpliSens® ARVI-screen-FRT PCR kit	1x10 ³

11.2 Specificity

AmpliSens® ARVI-screen-FRT PCR kit makes it possible to detect cDNA/DNA specific regions of ARVI causative agents listed above. The specificity of this kit was confirmed by investigation of the following reference strains: *human Respiratory Syncytial virus* (subgroup A, Long strain), *human*

Rhinoviruses (13, 15, 16, 17, 21, 26, and 29 types). The specificity of the kit was also proved during examination of clinical material with subsequent confirmation by sequencing the amplification products of the following pathogens: *human Respiratory Syncytial virus* (types A and B); *Parainfluenza virus-1-4*; *human Coronaviruses* OC43, E229, NL63, and HKU1; *human Adenoviruses* B, C, and E; *Metapneumoviruses A and B*; and *human Bocavirus*. It is also possible to discriminate between closely related enteroviruses in the reaction for rhinovirus RNA detection. The adenovirus detection reaction is not intended for typing because of possible interaction with closely related adenoviruses of other types.

Non-specific reactions between the components of the PCR kit and cDNA/DNA of other viral (*Influenza A and B viruses*, Urbani SARS-associated *Coronavirus* (Frankfurt), *Coronaviruses* causing feline infectious peritonitis (F1, F2, and F5) and swine transmissible gastroenteritis (TGEV1, TGEV8, and TGEV9), *Herpes viruses*, *Cytomegalovirus*, *Enteroviruses* (Echo9 and Echo30), and 60 samples of cerebrospinal fluid from meningitis patients containing *Enterovirus* RNA) and bacterial (*Streptococcus* spp., *Staphylococcus aureus*, *Mycoplasma influenza*, *Chlamydomphila pneumonia*, *Haemophilus influenza*, *Moraxella catarrhalis*, and *Legionella pneumophila*) agents that cause acute respiratory diseases as well as normal nasal and oropharyngeal human microflora and human cDNA/DNA are absent.

12. REFERENCES

- Handbook “Sampling, Transportation, and Storage of Clinical Material for PCR Diagnostics”, developed by Federal State Institution of Science Central Research Institute of Epidemiology of Federal Service for Surveillance on Consumers’ Rights Protection and Human Well-Being, Moscow, 2008.

13. QUALITY CONTROL

In compliance with Federal State Institution of Science “Central Research Institute of Epidemiology” ISO 13485-Certified Quality Management System, each lot of **AmpliSens® ARVI-screen-FRT** PCR kit has been tested against predetermined specifications to ensure consistent product quality.

³ Analytical sensitivity is expressed in genome equivalents of pathogen (GE) per 1 ml of sample.

14. EXPLANATION OF SYMBOLS



Manufacturer



Temperature limitation



Use by



Batch code



For *in Vitro* Diagnostic Use



Version



Catalogue number



Consult instructions for use



Contains sufficient for <n> tests



Caution, consult accompanying documents

NCA

Negative Control of Amplification

C-

Negative control of extraction

PCE

Positive Control of Extraction

C+

Positive control of amplification

IC

Internal Control STI-rec

RG

For working with Rotor-Gene 3000/6000 (Corbett Research)

Mx

For working with Mx3000P or Mx3005P (Stratagene)

iQ

For working with iCycler iQ and iQ5 (Bio-Rad)

List of Changes Made in the Instruction Manual

VER	Location of changes	Essence of changes
20.09.10		AmpliSens® ARVI-screen-FRT PCR kit variant FRT was deleted
20.10.10	Through the text	The names of infections are capitalized
	7.3.2 Amplification	Phrase «It is not allowed to perform « <i>Rhinovirus</i> » together with other tests from AmpliSens® ARVI-screen-FRT PCR kit when working with iCycler iQ and iQ5 instruments» is added
	Data analysis	Phrase before table 3 is changed
26.10.10	Stability and storage	Phrase about keeping away from light of PCR-mix-1-FL-F <i>hRSv</i> – <i>hMpv</i> , PCR-mix-1-FL-F <i>hPiv</i> 1/3, PCR-mix-1-FL-F <i>hPiv</i> 2/4, PCR-mix-1-FL-F <i>hCov</i> , PCR-mix-1-FL-F <i>hAdv</i> – <i>hBov</i> , and PCR-mix-1-FL-F <i>hRv</i> is added
13.11.10	Through the text	Catalogue numbers R-V57(iQ,Dt)-CE, R-V57(RG)-CE are added
	Content	Footnotes for Negative Control (C-) and Internal Control STI-rec (IC) are added